

Searching for cross-kingdom communication in plant-microbe interactions and implications for agriculture

Ena Šečić (Prof. Dr. Karl-Heinz Kogel)

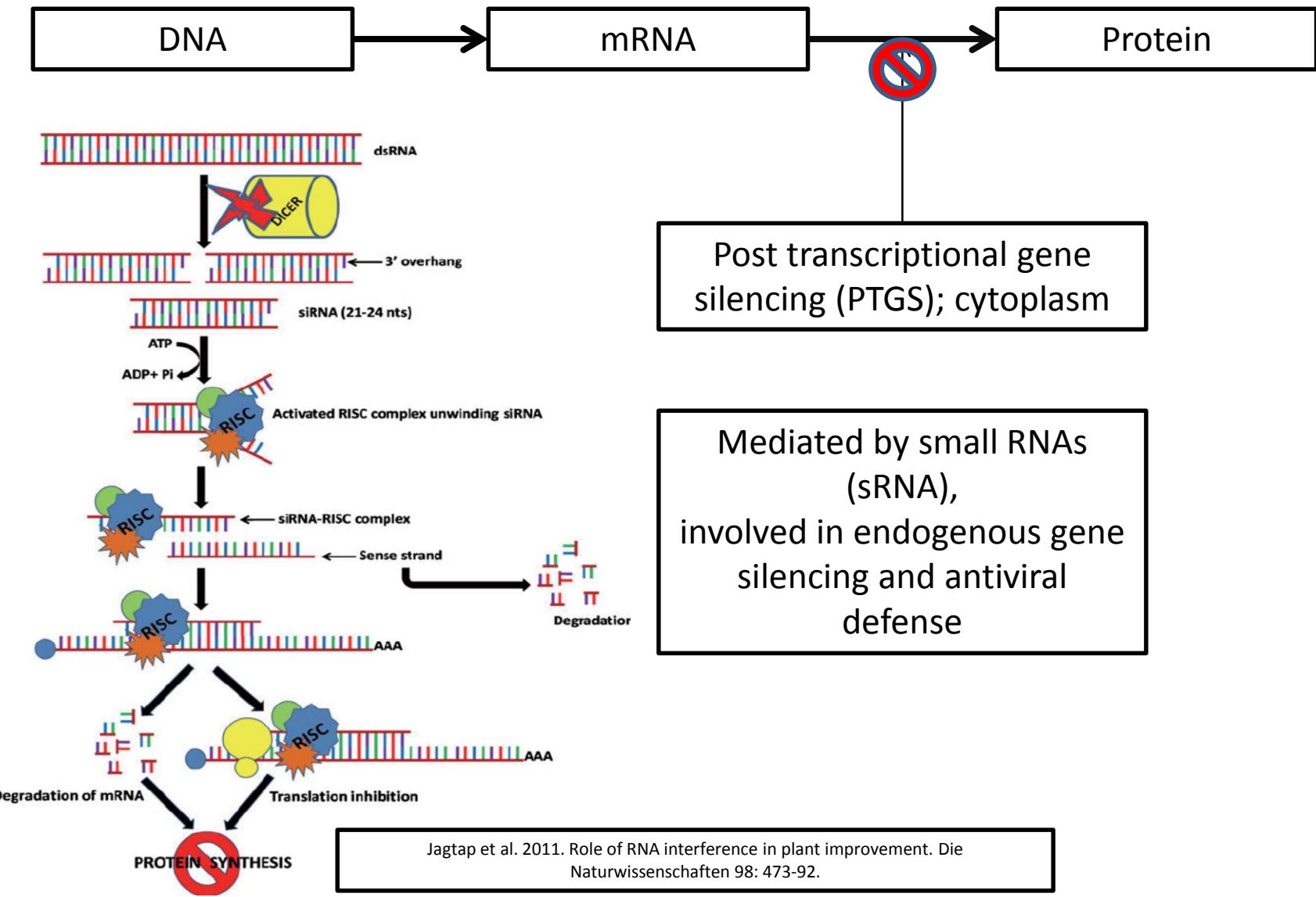
Institute for Phytopathology, Justus Liebig University
Giessen

Symposium

Modern Agriculture without Chemical
Pesticides?

September 03-04, 2018, Neustadt an der
Weinstraße

RNA Interference (RNAi)

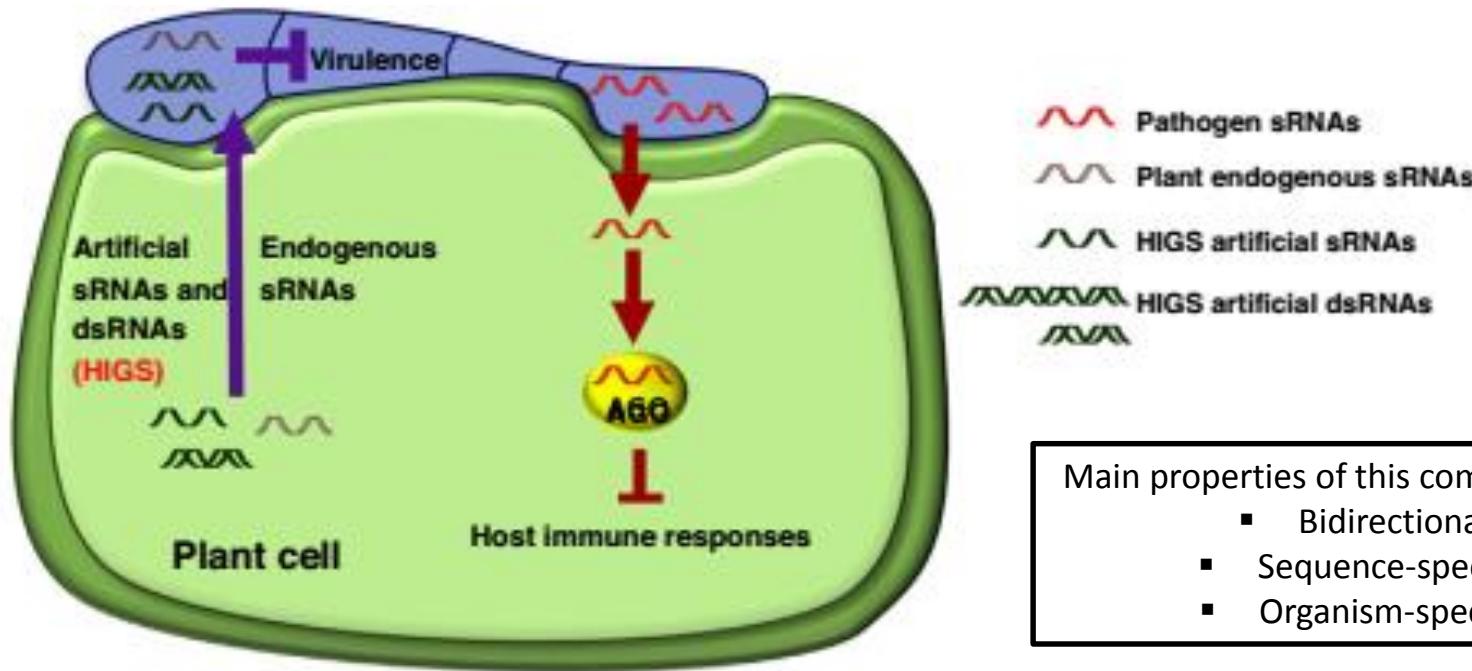


What is CROSS-KINGDOM
communication?
(in the context of plant-microbe
interactions)?

What is CROSS-KINGDOM communication? (in the context of plant-microbe interactions)?

- **Weiberg et al.** 2013. Fungal small RNAs suppress plant immunity by hijacking host RNA interference pathways. *Science* 342:118-123.
- **Wang M et al.** 2017a. Botrytis small RNA Bc-siR37 suppresses plant defense genes by cross-kingdom RNAi. *RNA Biol* 14:421-428.
- **Wang B et al.** 2017b. Puccinia striiformis f. sp. tritici microRNA-like RNA 1 (Pst-miR1), an important pathogenicity factor of Pst, impairs wheat resistance to Pst by suppressing the wheat pathogenesis-related 2 gene. *New Phytol* 215:338-350.
- **Zhang et al. 2016.** Cotton plants export microRNAs to inhibit virulence gene expression in a fungal pathogen. *Nat Plants* 2, doi:10.1038/nplants.2016.153.
- **Cai et al. 2018.** Cross-Kingdom RNAi - nature's blueprint for modern crop protection technology. *Current Opinion in Microbiology* 46:58-64, doi 10.1016/j.mib.2018.02.003.

Cross-kingdom RNAi in plant-microbe interactions

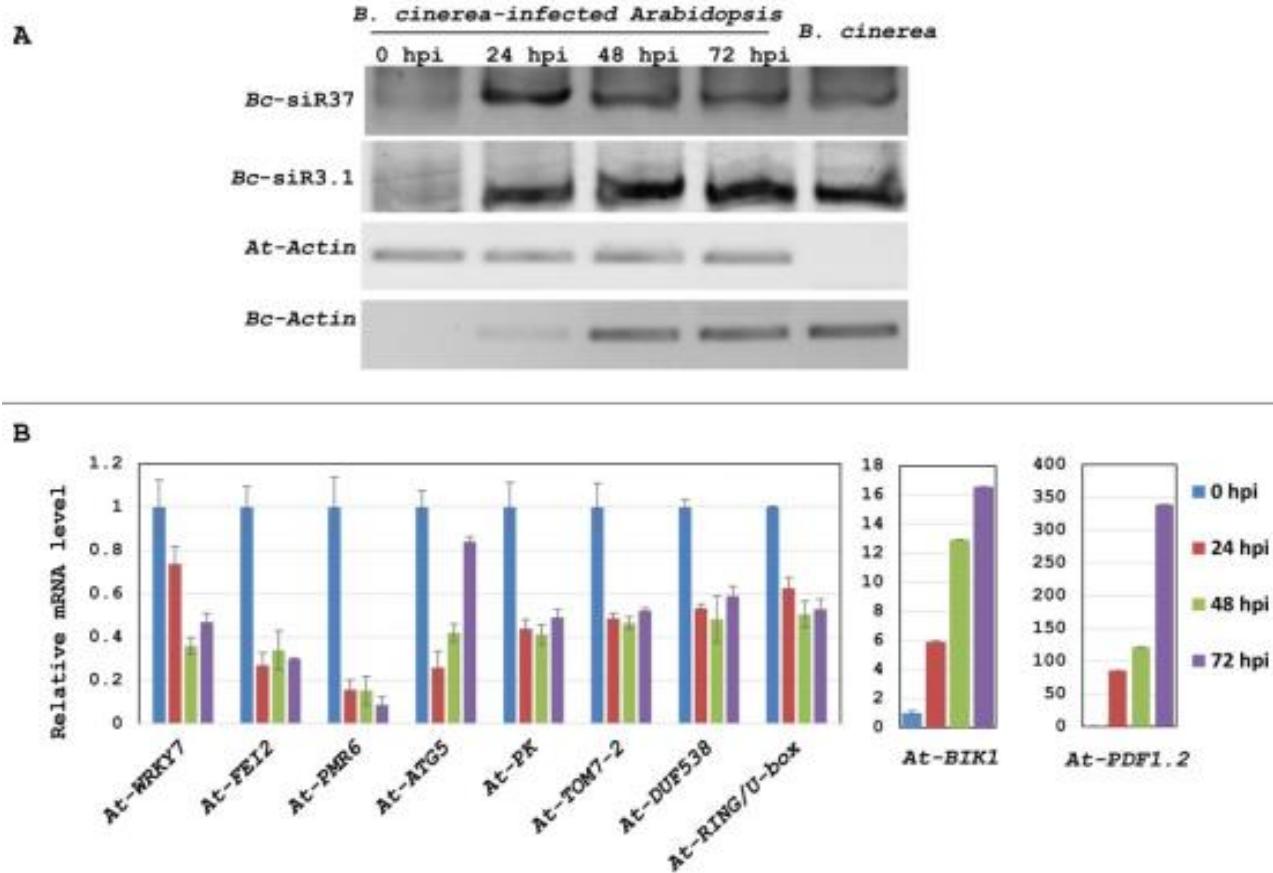


Main properties of this communication:

- Bidirectional
- Sequence-specific
- Organism-specific

Detected in plant-microbe and animal-microbe systems (even without a functional RNAi machinery)

Adjusted from: Cai et al. 2018. Cross-Kingdom RNAi - nature's blueprint for modern crop protection technology. DOI 10.1016/j.mib.2018.02.003.

Botrytis cinerea – Arabidopsis / tomato

Bc sRNA expressed in the Bc-Arabidopsis system and the corresponding target gene downregulation

From: Wang M et al. 2017a. Botrytis small RNA Bc-siR37 suppresses plant defense genes by cross-kingdom RNAi. RNA Biol 14:421-428.

Plant

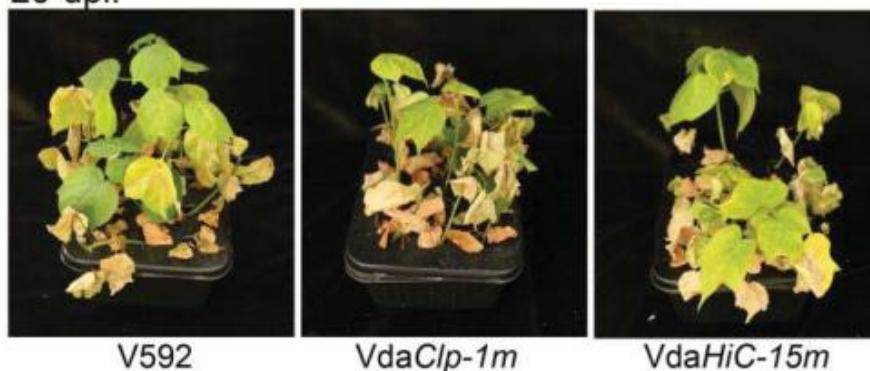
Pathogen / Microbe

Cotton - *Verticillium dahliae*

d 20-dpi.



f 20-dpi.



Mutant Vd strains with knock-out of mRNA targets (d) and with targets modified to be ck-RNA resistant (f) have resistant (d) and susceptible (f) responses on cotton plants

From: Zhang et al. 2016. Nat Plants 2, doi:10.1038/nplants.2016.153.

Plant

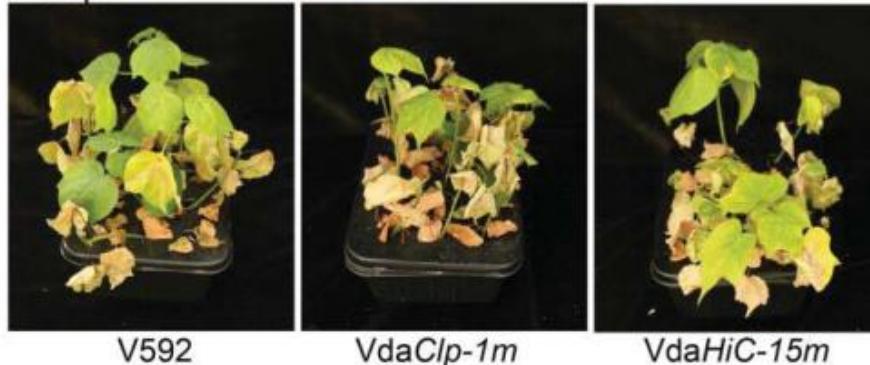
Pathogen Microbe

Cotton - *Verticillium dahliae*

d 20-dpi.



e 20-dpi.



Mutant Vd strains with knock-out of mRNA targets (d) and with targets modified to be ck-RNA resistant (e) have resistant (d) and susceptible (e) responses on cotton plants

From: Zhang et al. 2016. Nat Plants 2, doi:10.1038/nplants.2016.153.

Plant

Pathogen / Microbe



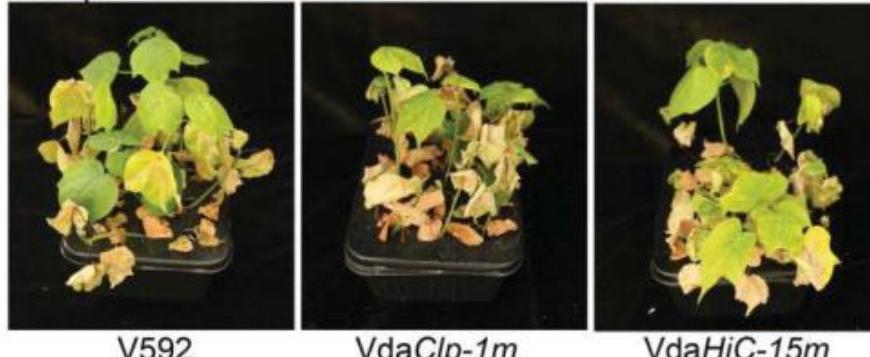
Cotton - *Verticillium dahliae*

d 20-dpi.



V592 Vda Δ c/p-1 Vda Δ hic-15

f 20-dpi.



V592

VdaC/p-1m

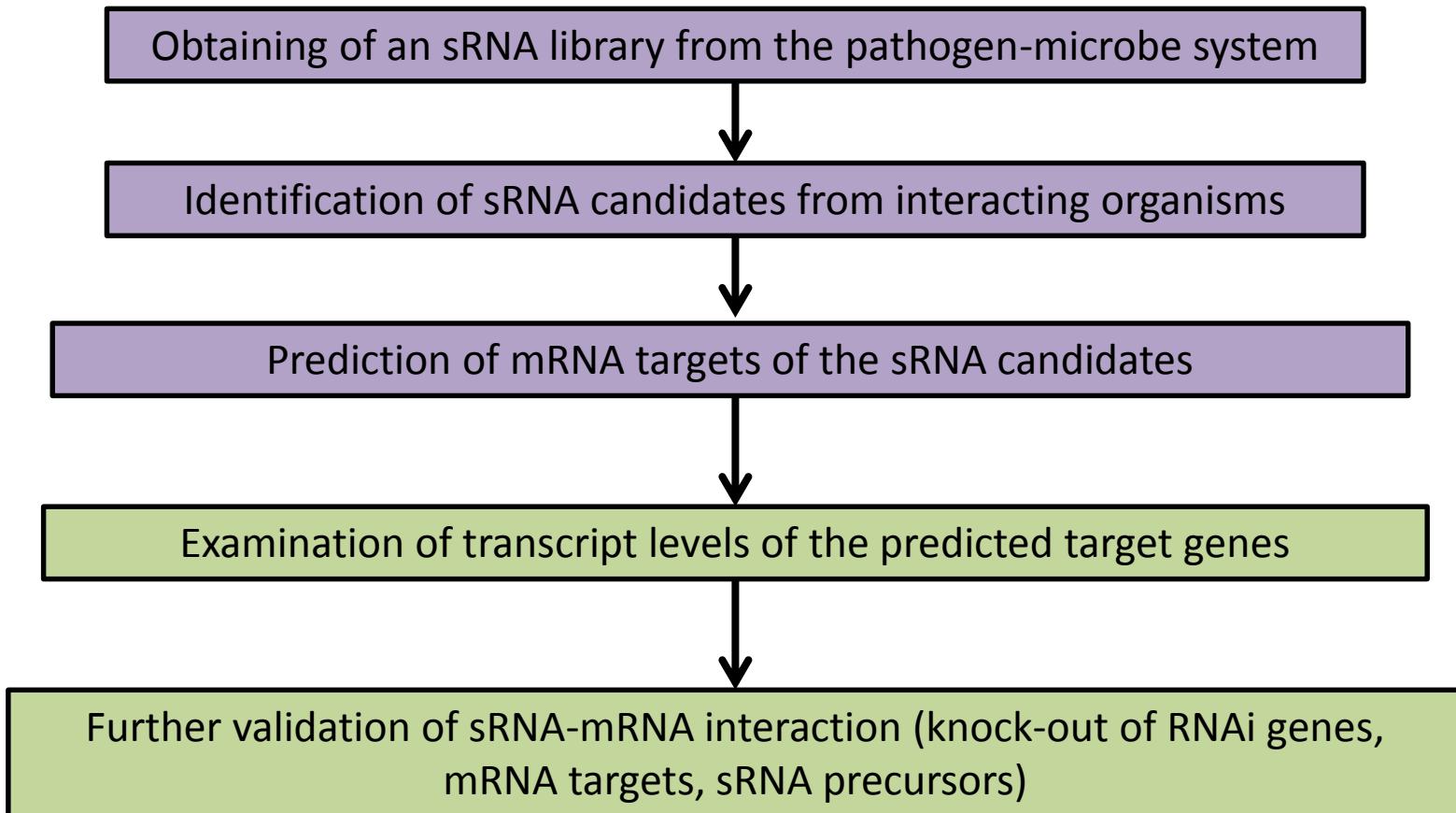
VdaHiC-15m

Mutant Vd strains with knock-out of mRNA targets (d) and with targets modified to be ck-RNA resistant (f) have resistant (d) and susceptible (f) reponses on cotton plants

From: Zhang et al. 2016. Nat Plants 2, doi:10.1038/nplants.2016.153.

Approaches to detect CROSS-KINGDOM communication

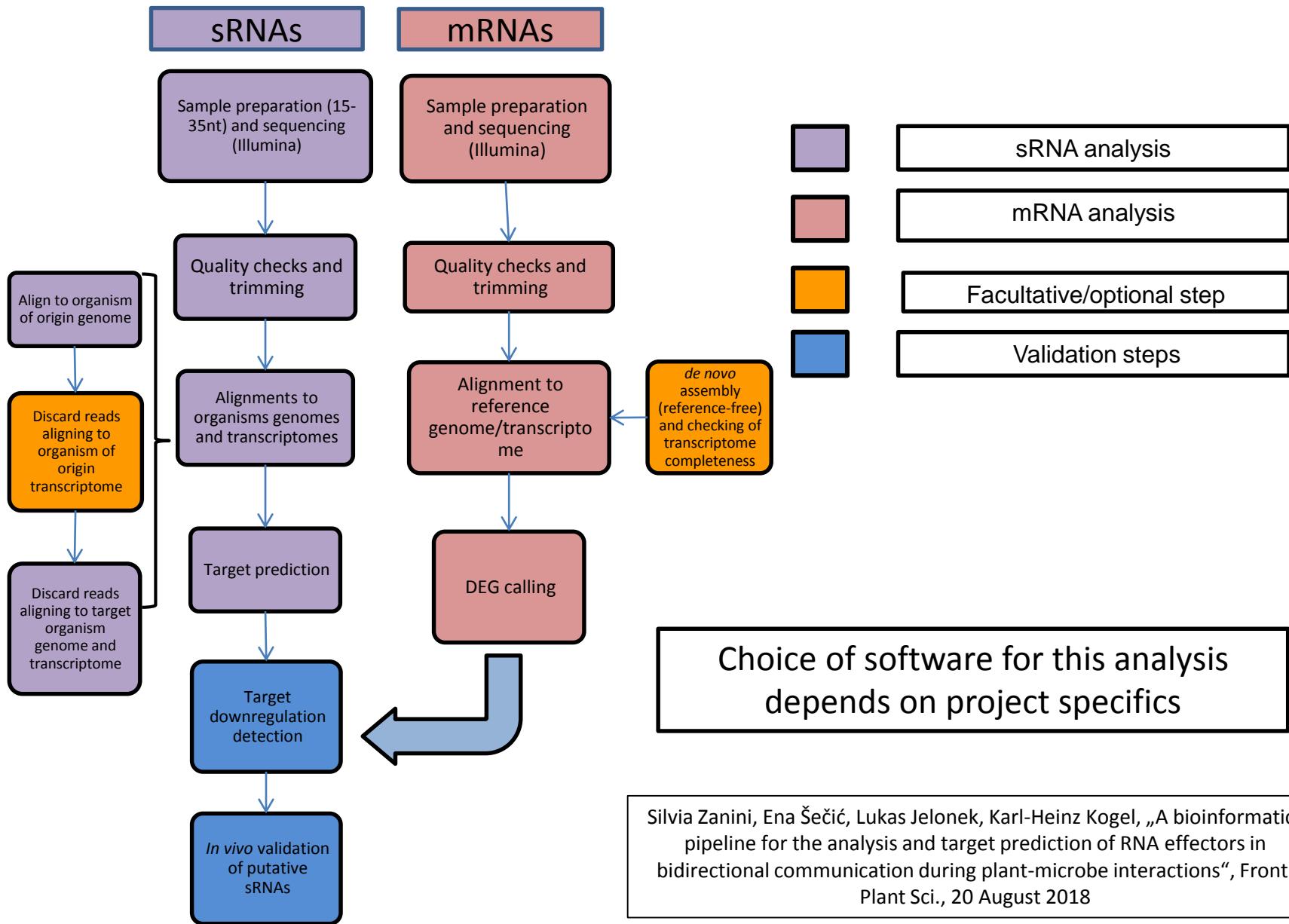
Cross-kingdom communication - identification pipeline



In silico steps



In vitro/in vivo steps

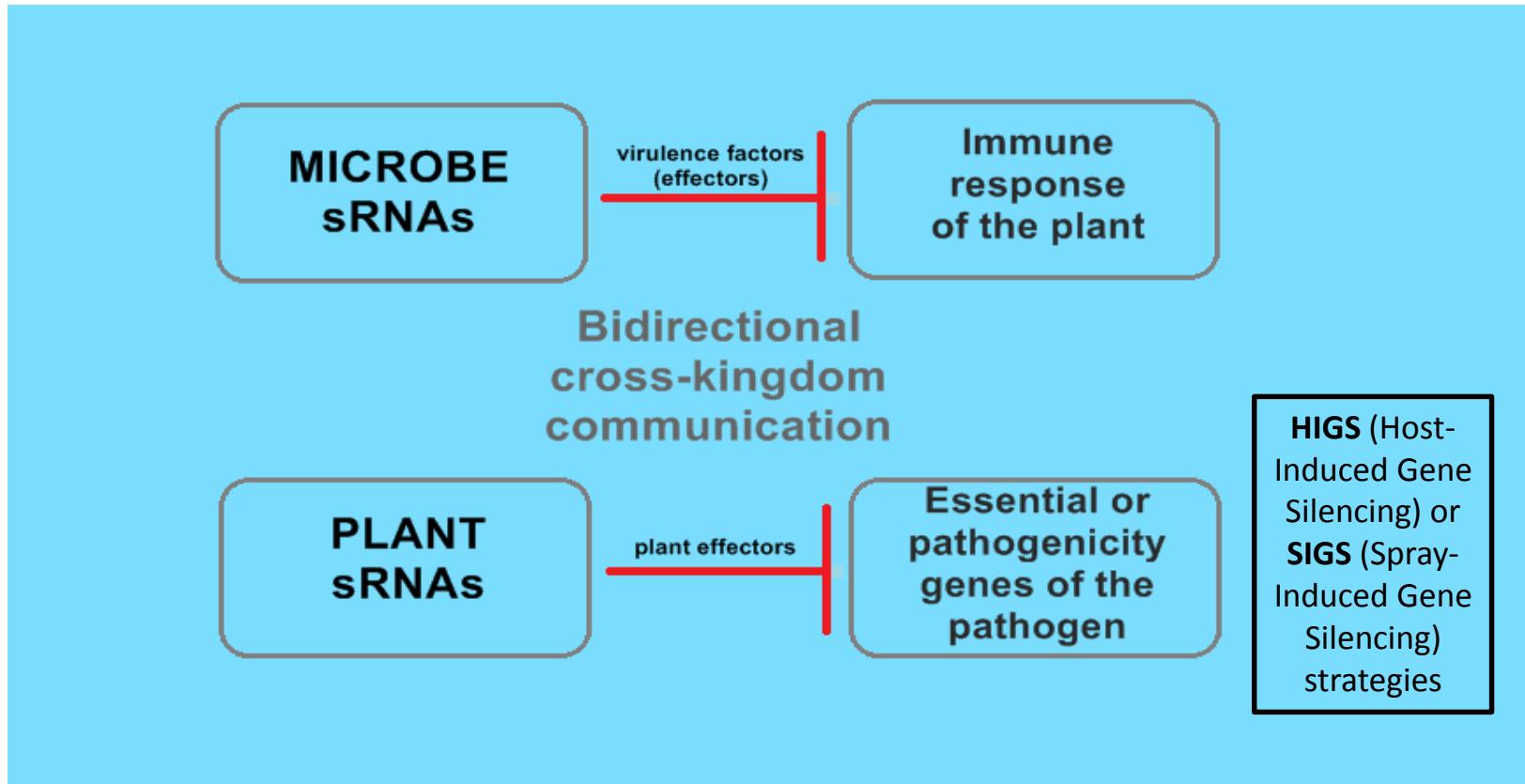


Fundamental research questions:

- Mechanistic background of cross-kingdom communication?
- How are sRNAs transported between organisms?
- Target conservation in microbes with different lifestyles?
- Target detection for design of plant protection strategies

Agronomic application strategies:

- HIGS and RNAs as fungicides / SIGS strategies



Application potential (ck-communication and RNAi protection strategies)

Chemical-based protection strategies to be avoided due to:

- Development of pathogen resistance
 - Soil/Water pollution
 - Pathogens difficult to control



**NOVEL PROTECTION STRATEGIES ARE CONSTANTLY
REQUIRED**



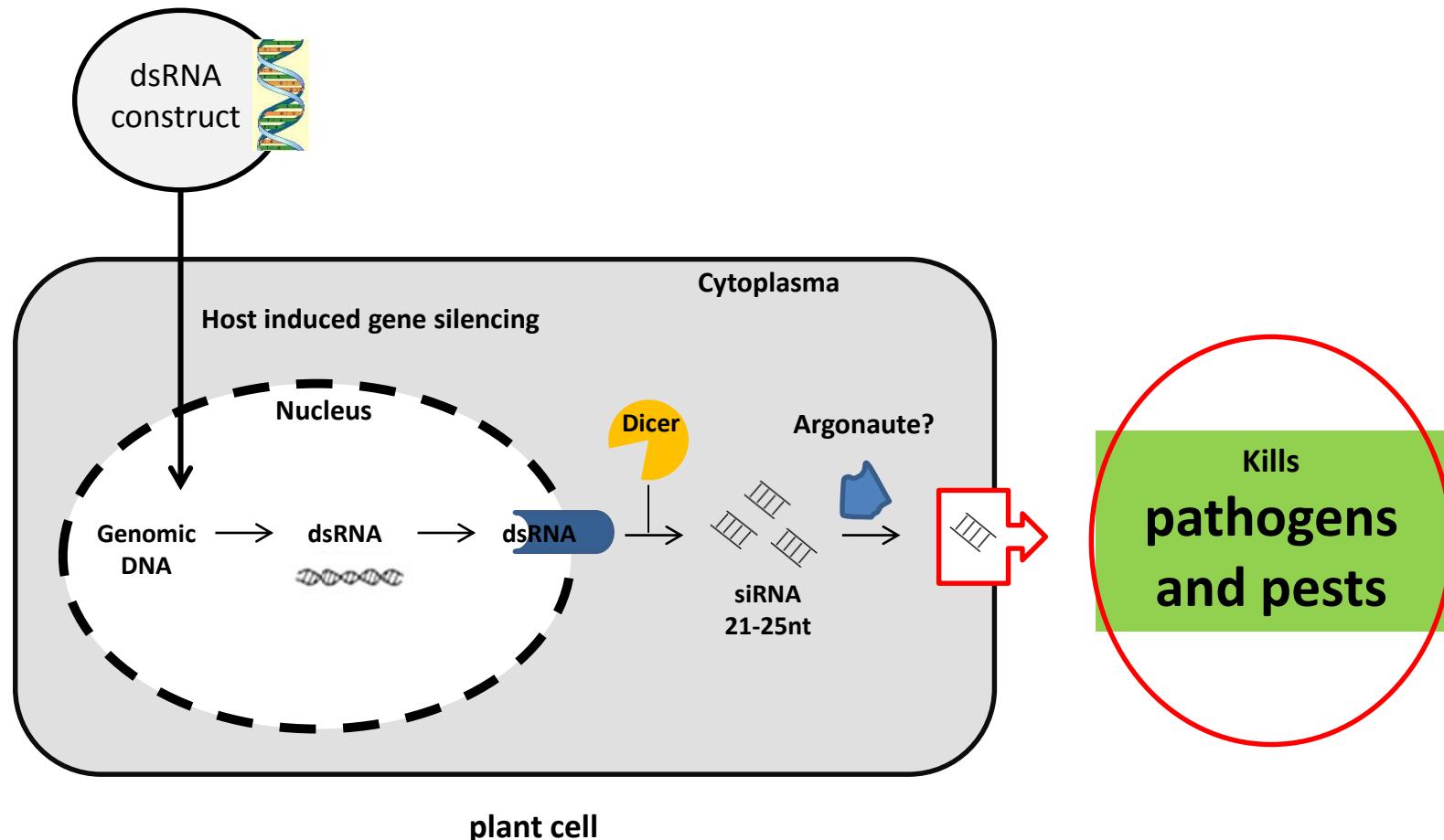
AGRICULTURAL APPLICATION OF dsRNA AS A SOLUTION

**Direct delivery via the crop plant (GMO)
(Host-Induced Gene Silencing, HIGS*)**

**Delivery via spray-application (RNA as fungicide)
(Spray-Induced Gene Silencing, SIGS)**

*Nowara et al. (2010) Plant Cell 22:3130-3141

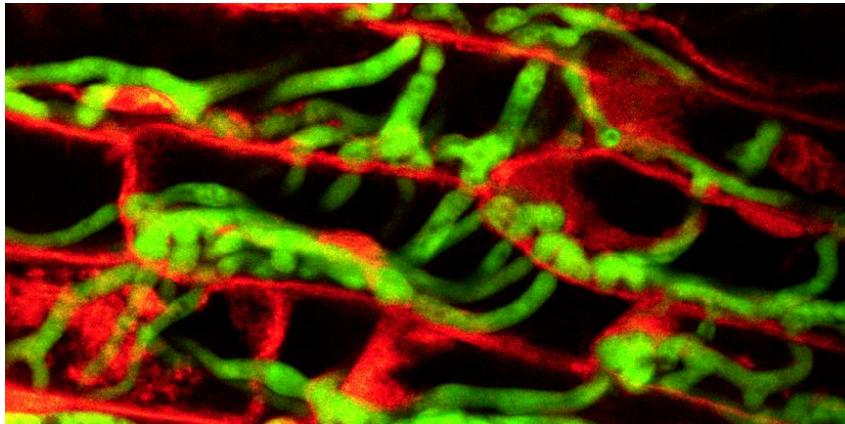
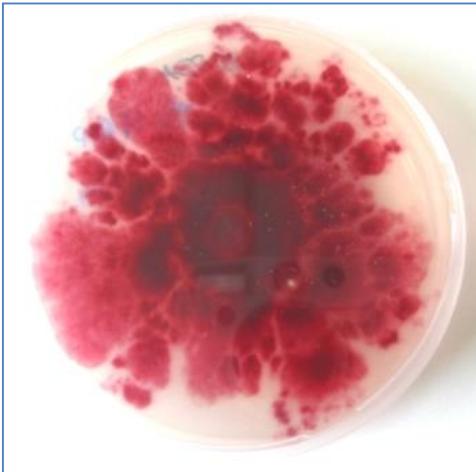
HIGS – A GMO plant protection strategy



Fusarium graminearum

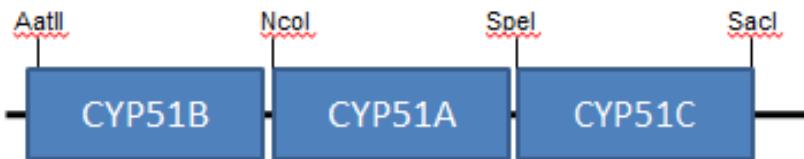


Macroconidia



Head blight disease

CYP3-dsRNA inhibits Fusarium *in vitro*

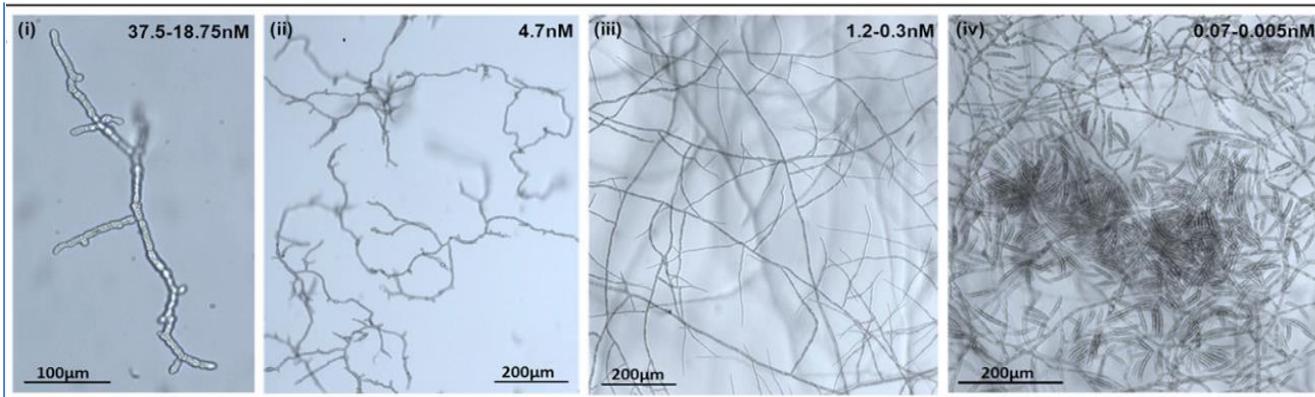


CYP3-dsRNA (791 nt long)



Fusarium graminearum

In vitro growth inhibition
by CYP3-dsRNA

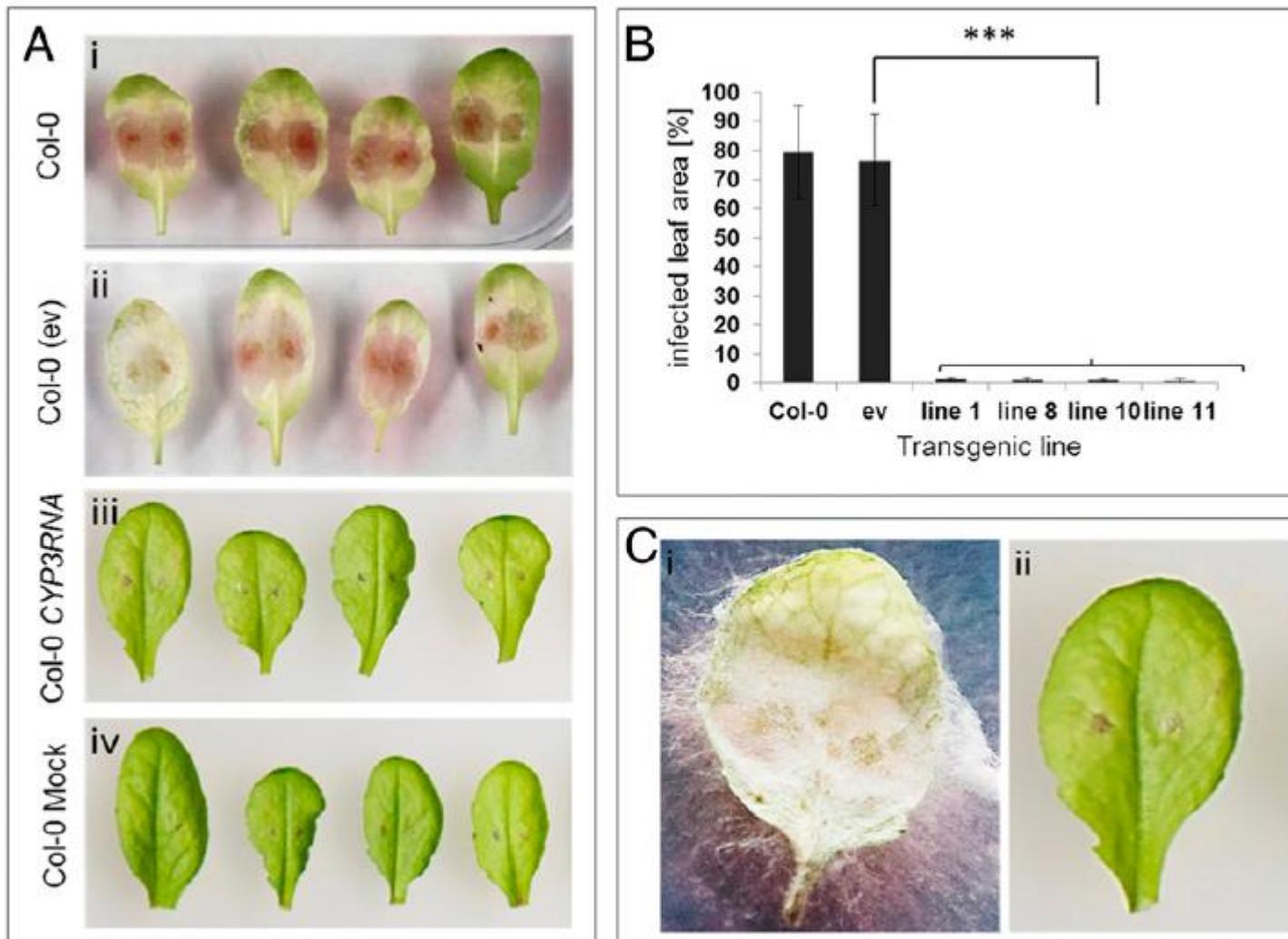


Amount of dsRNA

Koch et al. 2013, PNAS 110:19324

Koch et al. 2016, Plos Pathog. 10.1371/journal.ppat.1005901

CYP3-dsRNA inhibits Fusarium infection on transgenic Arabidopsis plants



Fusarium graminearum strain Fg-IFA65

12.9.2018

Secic Ena, Neustadt 2018

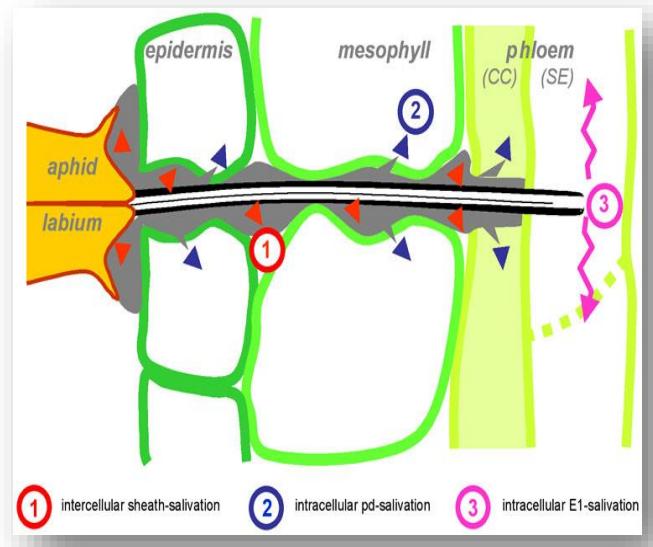
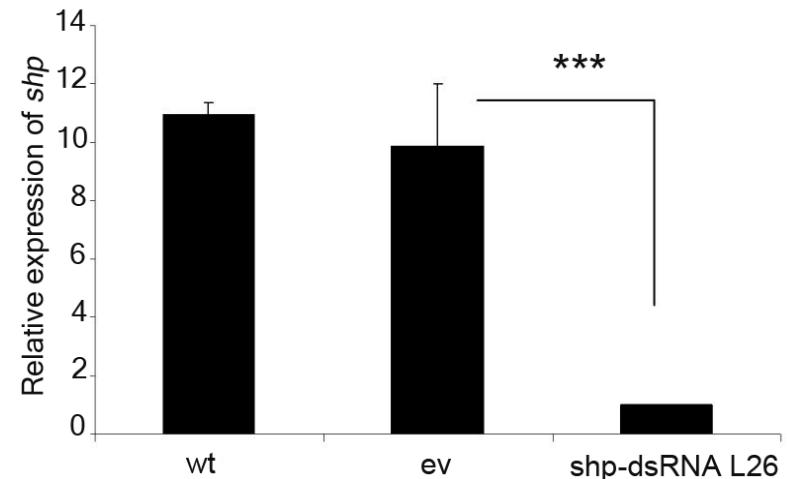
Koch et al. 2013, PNAS 110

20

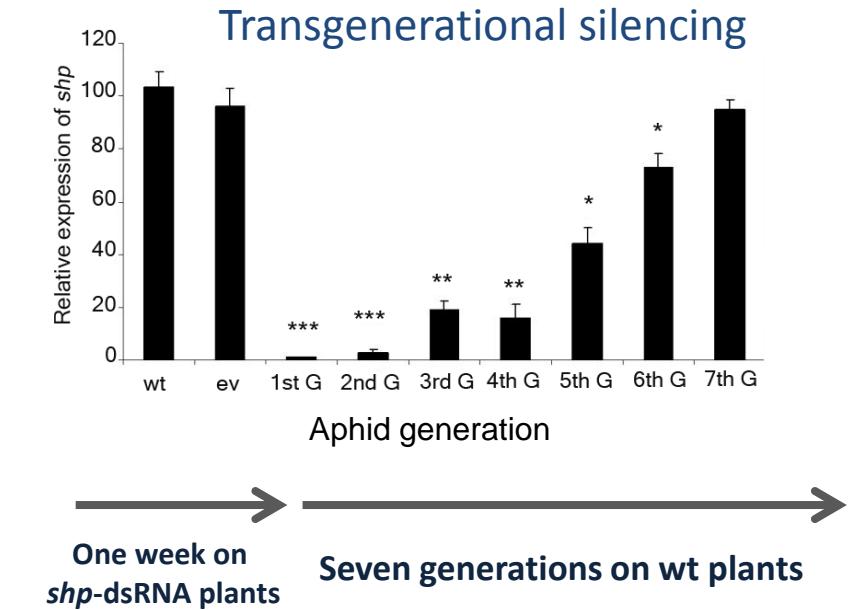
Aphid target „SHEATH PROTEIN“ (shp)



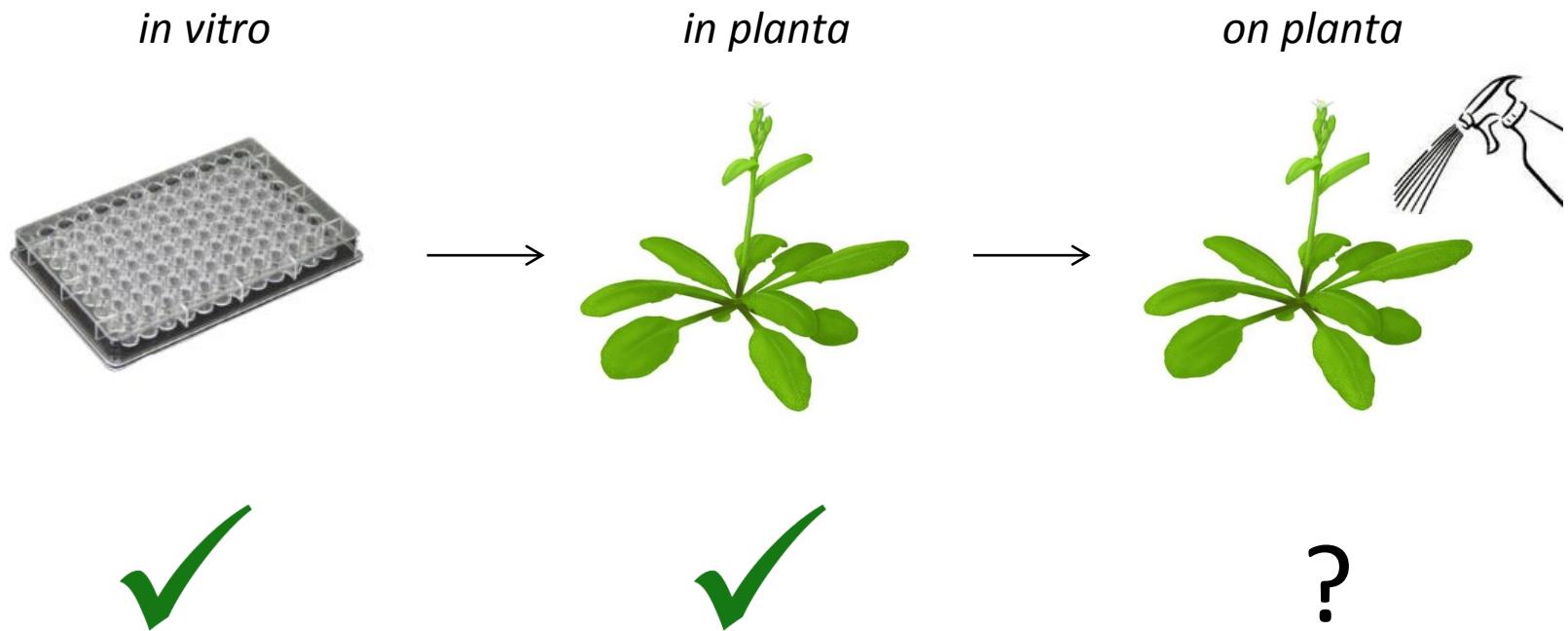
www.lastrefuge.co.uk

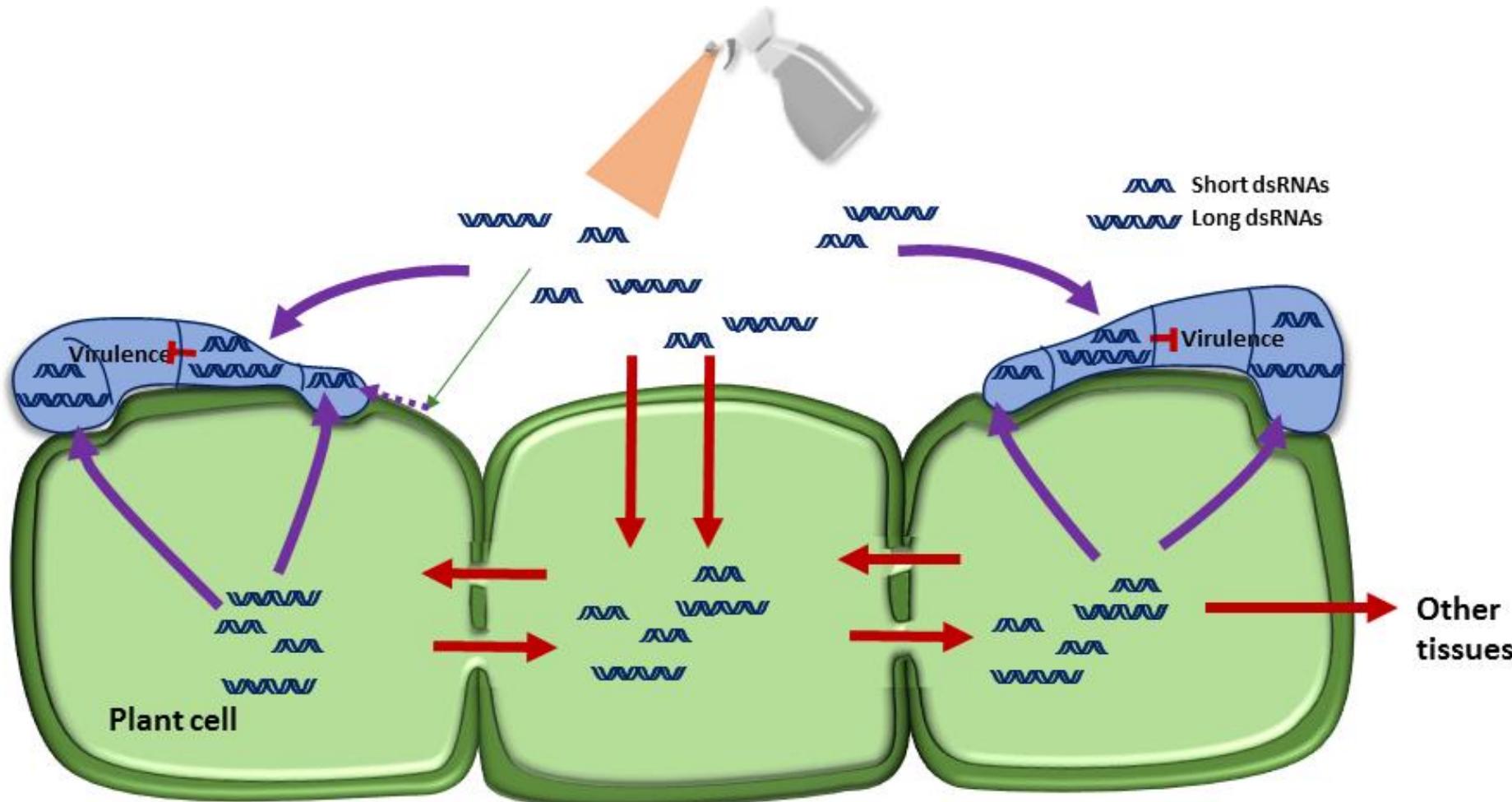


Abdellatif et al. 2014, Plant Biotech. J. 13:849



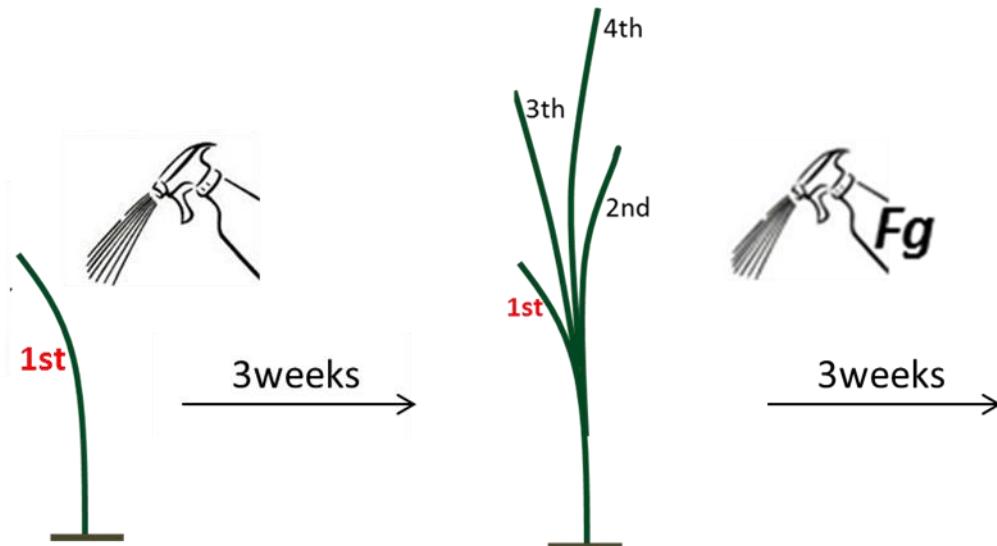
SIGS – dsRNA as a fungicide



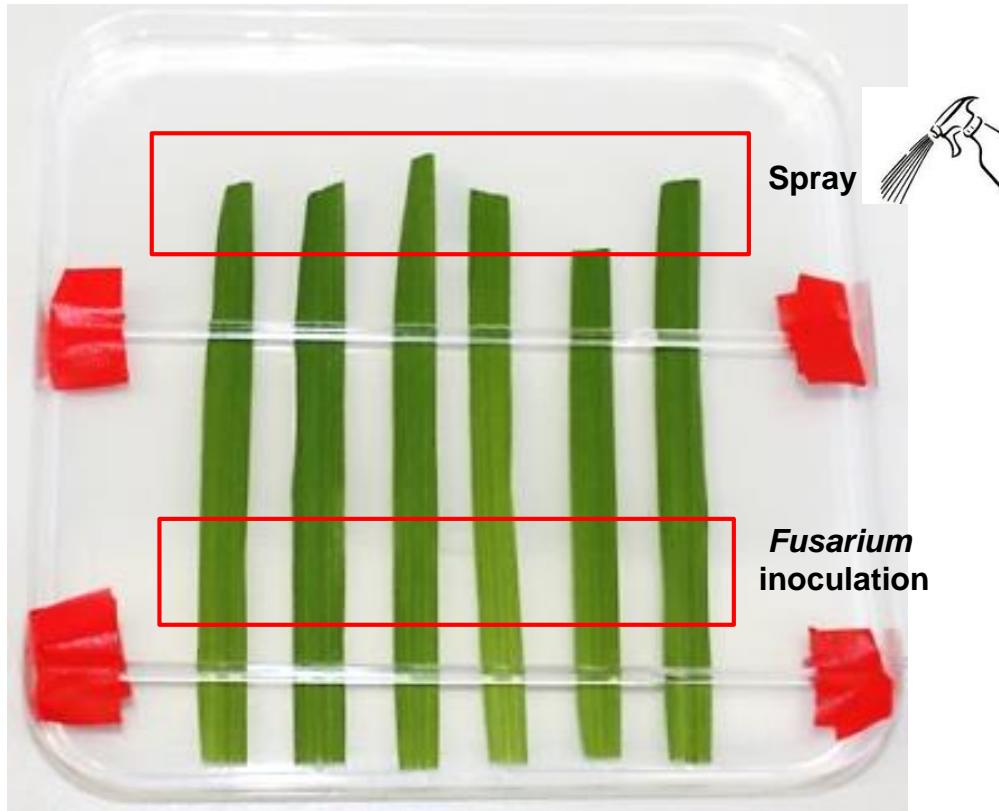


Adjusted from: Cai et al. 2018. Cross-Kingdom RNAi - nature's blueprint for modern crop protection technology. DOI 10.1016/j.mib.2018.02.003.

SIGS-mediated control of *F. graminearum* on whole barley plants



Systemic Setup: spray-induced gene silencing (SIGS)

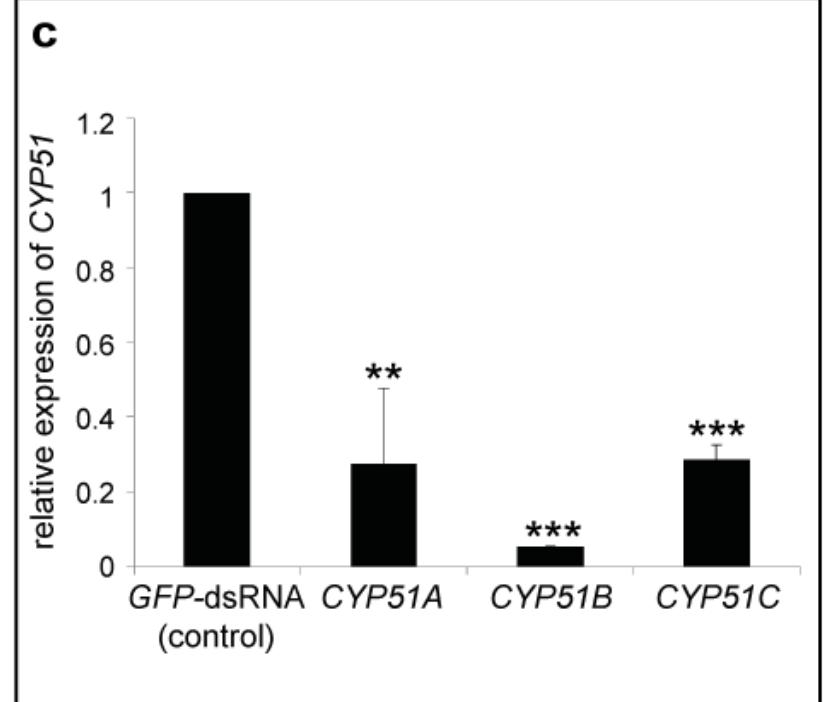
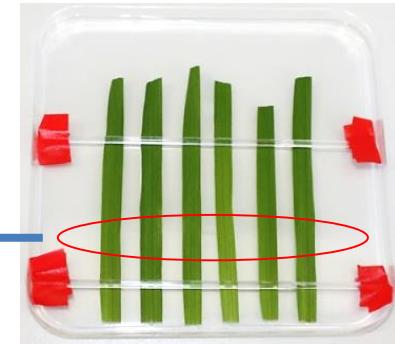
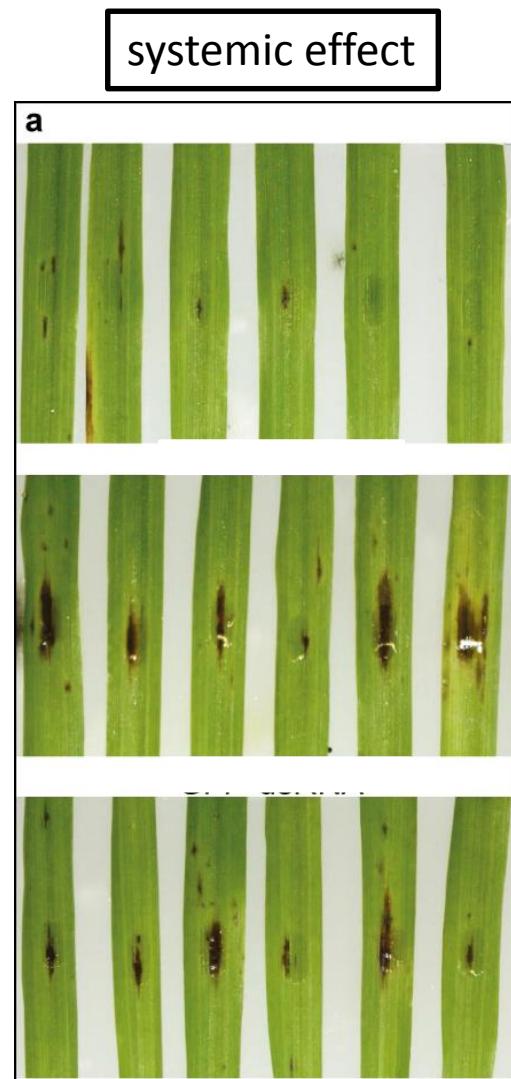


SIGS-mediated systemic control of *Fusarium graminearum*

CYP3-dsRNA

Buffer
control

GFP-dsRNA
control



RNAi-based crop protection: Application and potential drawbacks

- dsRNA design strategies
- Stability of dsRNA (formulation potential)
- Off-target prediction and precautions
- Emergence of resistant/tolerant microbes/pests?
- Accessibility for application and cost considerations

Off-target search

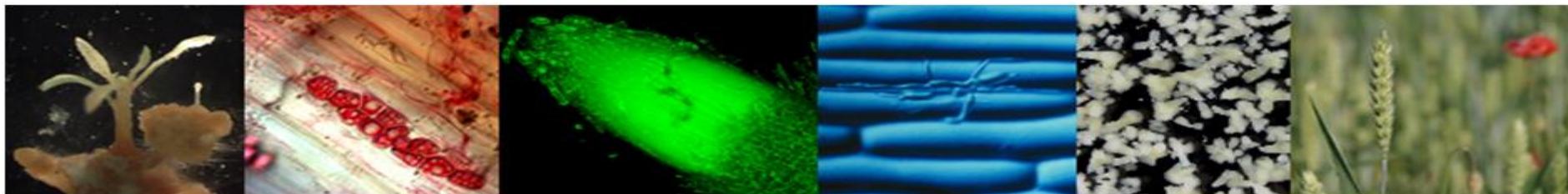
Table S1. Prediction of CYP3RNA off-target transcripts

Organism	Query name*			
	Gene	Description	All hits [†]	Efficient hits [‡]
<i>Fusarium graminearum</i>	FGSG_01000 [§]	CYP51B	200	46
	FGSG_04092 [§]	CYP51A	274	126
	FGSG_11024 [§]	CYP51C	218	95
<i>Arabidopsis thaliana</i>	AT2G17330	CYP51A1	0	0
	AT1G11680	CYP51A2	0	0
<i>Hordeum vulgare</i>	Published database (1)		0	0
<i>Hyaloperonospora arabidopsis</i>	Published database (2)		0	0
<i>Rhizophagus irregularis</i>	Published database (3)		0	0
<i>Piriformospora indica</i>	Published database (4)		0	0
<i>Homo sapiens</i>	Published database (5)		0	0
<i>Fusarium cerealis</i> isolate NRRL13721	JN416614 [¶]	CYP51A	190	83
<i>Fusarium austroamericanum</i> isolate NRRL28718	JN416607 [¶]	CYP51A	117	50
<i>Fusarium vorosii</i> isolate 67C1	JN416608 [¶]	CYP51A	116	50
<i>Fusarium acaciae-mearnsii</i> isolate NRRL26752	JN416603 [¶]	CYP51A	94	43

*Simulations were run using Si-Fi software (v3.1) for predicting off-targets prediction (<http://labtools.ipk-gatersleben.de>).

†Number of 21-mer siRNA sequences with perfect match to the query sequence.

‡Number of 21-mer siRNA sequences with perfect match to the query sequence that fulfill additional criteria for efficient RNAi (See Si-Fi software).



THANK YOU FOR ATTENTION!

